

**FINAL STUDY REPORT**STUDY TITLE

AOAC Use-Dilution Method

Test Organisms:*Pseudomonas aeruginosa* (ATCC 15442)*Staphylococcus aureus* (ATCC 6538)*Salmonella choleraesuis* (ATCC 10708)PROTOCOL NUMBER

SRC27022404.UD.1

PRODUCT IDENTITY

ACCEL TB

Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US

DATA REQUIREMENTS

U.S. EPA 40 CFR Part 158

"Data Requirements for Registration"

Pesticide Assessment Guidelines - Subdivision G, 91-2 (d)

AUTHOR

Sally Nada, B.S.

Study Director

STUDY COMPLETION DATE

June 21, 2004

PERFORMING LABORATORY

ATS Labs

1285 Corporate Center Drive, Suite 110

Eagan, MN 55121

SPONSOR

Virox Technologies

6705 Mill Creek Road Unit 4

Mississauga, Ontario L5N5M4

SPONSOR REPRESENTATIVE

Scientific & Regulatory Consultants, Inc.

102 1/2 South Chauncey Street

Columbia City, IN 46725-2306

PROJECT NUMBER

A02064

Page 1 of 16

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 2 of 16



STATEMENT OF NO DATA CONFIDENTIALITY CLAIMS

No claim of confidentiality is made for any information contained in this study on the basis of its falling within the scope of FIFRA Section 10 (d) (1) (A), (B), or (C).

Company: Virox Technologies

Company Agent: Sally Hayes

Agent for Virox Technologies
Title

Sally Hayes
Signature

Date: 09/20/04

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 3 of 16



GOOD LABORATORY PRACTICE STATEMENT

The study referenced in this report was conducted in compliance with U.S. Environmental Protection Agency Good Laboratory Practice (GLP) regulations set forth in 40 CFR Part 160.

The procedures not performed by or under the direction of ATS Labs are exempt from this Good Laboratory Practice Statement and include: characterization and stability of the compound(s).

Submitter: *Sally Hayes*
Sally Hayes, Agent for Virox Technologies

Date: 09/20/04

Sponsor: *Rhonda Jones*
Rhonda Jones, Agent for Virox Technologies

Date: 6-29-04

Study Director: *Sally Nada*
Sally Nada, B.S.

Date: 1/21/04

Project No. A02064
 Protocol Number: SRC27022404.UD.1

Virox Technologies
 Page 4 of 16



QUALITY ASSURANCE UNIT SUMMARY

Study: AOAC Use-Dilution Method

The objective of the Quality Assurance Unit is to monitor the conduct and reporting of non-clinical laboratory studies. This study has been performed under Good Laboratory Practice regulations (40 CFR Part 160) and in accordance to standard operating procedures and a standard protocol. The Quality Assurance Unit maintains copies of study protocols and standard operating procedures and has inspected this study on the date(s) listed below. Studies are inspected at time intervals to assure the integrity of the study.

Phase Inspected	Date	Study Director	Management
Critical Phase	April 7, 2004	April 7, 2004	May 3, 2004
Draft Report	April 30, 2004	April 30, 2004	
Final Report	June 21, 2004	June 21, 2004	June 21, 2004

The findings of these inspections have been reported to management and the Study Director.

Quality Assurance Auditor: Rachelle L. Evenson Date: 06/21/04

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 5 of 16



TABLE OF CONTENTS

Title Page 1

Statement of No Data Confidentiality Claims 2

Good Laboratory Practice Statement..... 3

Quality Assurance Unit Summary 4

Table of Contents 5

Study Personnel 6

General Study Information 7

Test Substance Identity 7

Study Dates 7

Objective 7

Summary of Results 8

Study Materials 8

Test Method 9

Study Controls 10

Study Acceptance Criteria 11

Protocol Changes 11

Test History 11

Data Analysis..... 11

Study Retention 12

References 12

Results 12

Analysis 13

Study Conclusion..... 13

Table 1: Control Results 14

Table 2: Carrier Population Control Results 14

Table 3: Neutralization Confirmation Control Results 15

Table 4: Test Results 16

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 6 of 16



STUDY PERSONNEL

STUDY DIRECTOR: Sally Nada, B.S.

Professional personnel involved:

- | | |
|----------------------------|--------------------------------------|
| Douglas G. Anderson, Ph.D. | - President |
| Karen M. Ramm, B.A. | - Technical Director |
| David Rottjakob, M.T. | - Microbiology Program Manager |
| Barbara Bailey, A.A. | - Microbiology Laboratory Supervisor |
| Sally Nada, B.S. | - Research Scientist I |
| Adam W. Pitt, B.S. | - Research Assistant II |
| Scott R. Steinagel, B.S. | - Research Assistant I |
| Matthew Sathe, B.S. | - Research Assistant I |
| Peter Toll, B.S. | - Research Assistant I |
| Lisa Slusser, B.S. | - Research Assistant I |

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 7 of 16



STUDY REPORT

GENERAL STUDY INFORMATION

Study Title: AOAC Use-Dilution Method

Project Number: A02064

Protocol Number: SRC27022404.UD.1

Sponsor: Virox Technologies
6705 Mill Creek Road Unit 4
Mississauga, Ontario L5N5M4

Sponsor Representative: Scientific & Regulatory Consultants, Inc.
102 1/2 South Chauncey Street
Columbia City, IN 46725-2306

Test Facility: ATS Labs
1285 Corporate Center Drive, Suite 110
Eagan, MN 55121

TEST SUBSTANCE IDENTITY

Test Substance Name: ACCEL TB

Lot/Batch(s): Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US

Test Substance Characterization

Test substance characterization as to content, stability, etc., (40 CFR, Part 160, Subpart F [160.105]) is the responsibility of the Sponsor.

STUDY DATES

Date Sample Received: March 11, 2004
Study Initiation Date: April 1, 2004
Experimental Start Date: April 7, 2004
Experimental End Date: April 26, 2004
Study Completion Date: June 21, 2004

OBJECTIVE

The objective of this study was to determine the efficacy of the Sponsor's product following the AOAC Use-Dilution Method in compliance with the U.S. Environmental Protection Agency requirements set forth in the Pesticide Assessment Guidelines.

Project No. A02064
 Protocol Number: SRC27022404.UJ.1

Virox Technologies
 Page 8 of 16



SUMMARY OF RESULTS

Test Substance: ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US)

Dilution: Ready to use (RTU)

Test Organisms: *Pseudomonas aeruginosa* (ATCC 15442)
Staphylococcus aureus (ATCC 6538)
Salmonella choleraesuis (ATCC 10708)

Exposure Time: One minute

Exposure Temperature: 20.0°C

Organic Soil Load: 5% fetal bovine serum

Efficacy Result: 3 lots of ACCEL TB demonstrated efficacy against *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Salmonella choleraesuis* as required by the U.S. EPA for disinfectant label claims.

STUDY MATERIALS

Test System/Growth Media

Test Organism	ATCC #	Growth Medium
<i>Pseudomonas aeruginosa</i>	15442	Nutrient Broth
<i>Staphylococcus aureus</i>	6538	Synthetic Broth
<i>Salmonella choleraesuis</i>	10708	Synthetic Broth

The microorganisms used in this study were obtained from the American Type Culture Collection, Manassas, Virginia.

Recovery Media

Neutralizing Subculture Medium: Lethen Broth with 1.0% Sodium Thiosulfate (Primary)
 Lethen Broth with 0.07% Lecithin & 0.5% Tween 80 (Secondary)

Agar Plate Medium: Tryptic Soy Agar with 5% Sheep Blood (BAP)

Reagents

Organic Soil Load Description: 5% fetal bovine serum (FBS)

Carriers

Stainless steel penicylinders were pre-soaked overnight in 1.0 N NaOH, washed in water until rinse water was neutral to phenolphthalein, and autoclaved in 0.1% asparagine.

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 9 of 16



TEST METHOD

Preparation of Test Substance

The test substance was ready to use (RTU), as received from the Sponsor. The test substance was homogenous as determined by visual observation.

Ten (10) mL aliquots of the test substance at the concentration(s) under test were transferred to sterile 25 x 150 mm tubes, placed in a 20±1°C water bath and allowed to equilibrate for ≥ 10 minutes.

Preparation of Test Organism

From a stock slant, an initial tube of culture broth was inoculated. From this initial broth suspension a minimum of three daily transfers was performed on consecutive days prior to use in testing procedure. For each test organism, the appropriate growth medium was subcultured using a daily transfer (more than 3, but less than 30 transfers) of the test organism.

A 48-54 hour broth culture incubated at 35-37°C was prepared. On the day of use, the pellicle was aspirated from the *Pseudomonas aeruginosa* culture. The test cultures were thoroughly mixed and allowed to stand for ≥10 minutes prior to use.

Per Sponsor's request, the organisms were standardized to meet a $1 \times 10^4 - 5 \times 10^6$ CFU/carrier specification. Based upon carrier counts typically achieved with undiluted broth cultures, *S. aureus* was diluted 1:50, *S. choleraesuis* was diluted 1:5 on 4/7/04 and 1:20 on 4/21/04, and *P. aeruginosa* was diluted 1:100. All cultures were diluted using Butterfield's Buffer. The diluted cultures were thoroughly mixed to assure homogeneity, and used to contaminate carriers. The carrier control results demonstrated that the CFU/carrier exceeded the EPA minimum standard of 1×10^4 CFU/carrier for all organisms and fell within the range specified by the Sponsor.

Addition of Organic Soil Load

On 4/7/04 a 9.4 mL aliquot of FBS was added to 178.6 mL of each broth culture to yield a 5% fetal bovine serum soil load. On 4/21/04 a 9.5 mL aliquot of FBS was added to 180.5 mL of *S. choleraesuis* broth culture to yield a 5% FBS soil load.

Contamination of Carriers

Sterile penicylinders were immersed for 15 minutes in a 48-54 hour old broth culture of the test organism, at a ratio of 1 carrier per 1.0 mL broth. The penicylinders were then dried on filter paper in a sterile petri dish at 35-37°C for 40 minutes and at 48.2% - 62.1% humidity.

Exposure Conditions

For each test substance, 60 contaminated and dried carriers were individually transferred by hook needle at staggered intervals to individual tubes containing 10 mL of the test substance at the requested dilution and exposed for one minute at 20.0°C.

Test System Recovery

Following exposure, each exposed carrier was then transferred by hook needle at identical staggered intervals to 10 mL of Letheen Broth with 1.0% Sodium Thiosulfate. Carriers were transferred from primary subculture tubes into individual secondary subculture tubes containing 10 mL of Letheen Broth with 0.07% Lecithin and 0.5% Tween 80 between 30-60 minutes following the first transfer.

Project No. A02064

Virox Technologies



Protocol Number: SRC27022404.UD.1

Page 10 of 16

Incubation and Observation

The neutralized subcultures were incubated for 48±4 hours at 35-37°C. Following incubation, the subcultures were examined for the presence or absence of visible growth.

Representative neutralized subcultures showing growth were subcultured, stained and/or biochemically assayed to confirm or rule out the presence of the test organism.

STUDY CONTROLS

Purity Control

A "streak plate for isolation" was performed on the organism culture and following incubation examined in order to confirm the presence of a pure culture. The acceptance criterion for this study control is a pure culture demonstrating colony morphology typical of the test organism.

Organic Soil Sterility Control

The serum used for soil load was cultured, incubated, and observed for lack of growth. The acceptance criterion for this study control is lack of growth.

Carrier Sterility Control

A representative uninoculated carrier was added to the subculture medium. The subculture medium containing the carrier was incubated and examined for growth. The acceptance criterion for this study control is lack of growth.

Neutralizing Subculture Medium Sterility Control

A representative sample of uninoculated neutralizing subculture medium was incubated and observed. The acceptance criterion for this study control is lack of growth.

Viability Control

A representative inoculated carrier was added to the subculture medium. The subculture medium containing the carrier was incubated and examined for growth. The acceptance criterion for this study control is growth.

Neutralization Confirmation Control

The neutralization of the test substance was confirmed by exposing sterile carriers (representing not less than 10% of the total number of test carriers) to the test substance and transferring them to primary subculture tubes containing 10 mL of neutralizing subculture medium. Carriers were then transferred from primary subculture tubes into individual secondary subculture tubes between 30-60 minutes following the primary transfer. The subculture tubes containing the exposed carriers were inoculated with ≤100 CFU of the test organism, incubated under test conditions and observed for the presence of growth. This control was performed with multiple replicates using different dilutions of the test organism. A standardized spread plate procedure was run concurrently in order to enumerate the number of CFU actually added. The control result was reported using data from the most appropriate dilution.

The acceptance criterion for this study control is growth after inoculation with ≤100 CFU.

Carrier Population Control

Inoculated carriers were added at a ratio of 1 carrier to 10 mL neutralizing broth and vortex mixed. Appropriate serial ten-fold dilutions were prepared and aliquots were spread plated on agar plate medium, and incubated. Following incubation, the resulting colonies were enumerated and the CFU/carrier calculated. The acceptance criterion for this study control is an average count of between 1×10^4 and 5×10^6 CFU/carrier per Sponsor's request.

Project No. A02064
Protocol Number: SRC27022404.UJ.1.

Virox Technologies
Page 11 of 16

ATS LABS

STUDY ACCEPTANCE CRITERIA

Test Substance Performance Criteria

The EPA efficacy performance requirements for label claims state that the disinfectant must kill the microorganisms on 59 out of the 60 inoculated carriers.

Control Acceptance Criteria

The study controls must perform according to the criteria detailed in the study controls description section.

PROTOCOL CHANGES

Protocol Amendments:

- 1) This protocol was amended to rectify the incorrect test substance expiration date for Lot 1-3635-REG-US listed in the protocol. The protocol was amended to change the expiration date from February 14, 2005 to February 12, 2005.
- 2) This protocol was amended per Sponsor's request to repeat all three lots against *Salmonella choleraesuis* (ATCC 10708) to test for the possibility of false positive results.
- 3) This protocol is amended per Sponsor's request to correct the test substance lot numbers listed in the protocol. The test substance name should read Lot 1-3635-REG-US (60 day old lot), Lot 2-3646-REG-US, and Lot 3-3647-REG-US instead of Lot 3635-REG-US (60 day old), Lot 3646-REG-US and Lot 3647-REG-US.

Protocol Deviations:

No protocol deviations occurred during this study.

TEST HISTORY

Testing was repeated against *Salmonella choleraesuis* (ATCC 10708) on 4/21/04 to test for the possibility of false positive results. All three lots were re-tested based on the EPA's Clarification Policy and correspondence with Nancy Whyte on 10/22/03.

DATA ANALYSIS

Calculations

Carrier Population Control Calculation:

$$\text{CFU/carrier} = \frac{(\text{average number colonies/plate @ dilution}) \times (\text{dilution factor}) \times (\text{volume neutralizer})}{(\text{number of carriers tested}) \times (\text{volume plated})}$$

The carrier population was calculated and reported using data from the most appropriate dilution(s).

Statistical Analysis

None used.

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 12 of 16



STUDY RETENTION

Record Retention

All of the original raw data developed exclusively for this study shall be archived at ATS Labs, 1285 Corporate Center Drive, Suite 110, Eagan, MN 55121. The original data includes, but is not limited to, the following:

1. Certified copy of final study report.
2. Original signed protocol.
3. Any protocol amendments.
4. All handwritten raw data for control and test substances including, but not limited to notebooks, data forms and calculations.
5. All measured data used in formulating the final report.
6. Memoranda, specifications, and other study specific correspondence relating to interpretation and evaluation of data, other than those documents contained in the final study report.
7. Study-specific SOP deviations made during the study.

Test Substance Retention

The test substance will be discarded following study completion per Sponsor approved protocol. It is the responsibility of the Sponsor to retain a sample of the test material.

REFERENCES

1. Association of Official Analytical Chemists (AOAC), 1990. Use-Dilution Tests, p. 135-137. *In* Official Methods of Analysis of the AOAC, Fifteenth Edition.
2. Association of Official Analytical Chemists (AOAC), 1990. Germicidal and Detergent Sanitizing Action of Disinfectants, p. 139 [Preparation of Synthetic Hard Water]. *In* Official Methods of Analysis of the AOAC, Fifteenth Edition.
3. U.S. Environmental Protection Agency, Registration Division, Office of Pesticide Programs, 1982. Efficacy Data Requirements, Disinfectants for Use on Hard Surfaces, DIS/TSS-1.
4. U.S. Environmental Protection Agency, Registration Division, Office of Pesticide Programs, 1979. Efficacy Data Requirements, Supplemental Recommendations, DIS/TSS-2.
5. U.S. Environmental Protection Agency, Registration Division, Office of Pesticide Programs, 1982. Subseries 91A: Public Health Uses. *In* Pesticide Assessment Guidelines – Subdivision G (Product Performance).

RESULTS

For Control and Neutralization Results, see Tables 1-3.

All data measurements/controls including the culture purity, viability, organic soil sterility, neutralizing subculture medium sterility, carrier sterility, neutralization confirmation, and carrier population were within acceptance criteria.

For Test Results, see Table 4.

Project No. A02064
Protocol Number: SRC27022404.UD.1

Virox Technologies
Page 13 of 16

ATS LABS

ANALYSIS

ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), ready to use, demonstrated growth of *Pseudomonas aeruginosa* (ATCC 15442) in 0, 0, 0, respectively, of the 60 primary subculture tubes and growth in 0, 1, 0, respectively, of the 60 secondary subculture tubes following a one minute exposure period in the presence of a 5% fetal bovine serum organic soil load at 20.0°C.

ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), ready to use, demonstrated growth of *Staphylococcus aureus* (ATCC 6538) in 0, 0, 0, respectively, of the 60 primary subculture tubes and growth in 1, 0, 0, respectively, of the 60 secondary subculture tubes following a one minute exposure period in the presence of a 5% fetal bovine serum organic soil load at 20.0°C.

ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), ready to use, demonstrated growth of *Salmonella choleraesuis* (ATCC 10708) in 1, 0, 1, respectively, of the 60 primary subculture tubes and growth in 3, 3, 1, respectively, of the 60 secondary subculture tubes following a one minute exposure period in the presence of a 5% fetal bovine serum organic soil load at 20.0°C. To test for the possibility of false positive results, repeat testing under the same conditions of ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), demonstrated growth of *Salmonella choleraesuis* (ATCC 10708) in 0, 1, 0, respectively, of the 60 primary subculture tubes and growth in 1, 0, 1, respectively, of the 60 secondary subculture tubes.

STUDY CONCLUSION

Under the conditions of this investigation, in the presence of a 5% fetal bovine serum organic soil load, ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), ready to use, demonstrated efficacy against *Pseudomonas aeruginosa* after a one minute contact at 20.0°C as required by the U.S. EPA for disinfectant label claims.

Under the conditions of this investigation, in the presence of a 5% fetal bovine serum organic soil load, ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), ready to use, demonstrated efficacy against *Staphylococcus aureus* after a one minute contact at 20.0°C as required by the U.S. EPA for disinfectant label claims.

Under the conditions of this investigation, in the presence of a 5% fetal bovine serum organic soil load, ACCEL TB (Lot 1-3635-REG-US (60 day old), Lot 2-3646-REG-US and Lot 3-3647-REG-US), ready to use, demonstrated efficacy against *Salmonella choleraesuis* after a one minute contact at 20.0°C as required by the U.S. EPA for disinfectant label claims.

In the opinion of the Study Director, there were no circumstances that may have adversely affected the quality or integrity of the data.

The use of the ATS Labs name, logo or any other representation of ATS Labs without the written approval of ATS Labs is prohibited. In addition, ATS Labs may not be referred to in any form of promotional materials, press releases, advertising or similar materials (whether by print, broadcast, communication or electronic means) without the express written permission of ATS Labs.

Project No. A02064

Virox Technologies



Protocol Number: SRC27022404.UD.1

Page 14 of 16

TABLE 1: CONTROL RESULTS

The following results from controls confirmed study validity:

Type of Control	Results			
	<i>Pseudomonas aeruginosa</i> (ATCC 15442)	<i>Staphylococcus aureus</i> (ATCC 6538)	<i>Salmonella choleraesuis</i> (ATCC 10708)	
	4/7/04	4/7/04	4/7/04	4/21/04
Purity Control	Pure	Pure	Pure	Pure
Viability Control	Growth	Growth	Growth	Growth
Organic Soil Sterility Control (3 Vials utilized on 4/7/04 testing)	No Growth; No Growth; No Growth			No Growth
Neutralizing Subculture Medium Sterility Control (2 lots utilized)	No Growth; No Growth			No Growth; No Growth
Carrier Sterility Control	No Growth			No Growth

TABLE 2: CARRIER POPULATION CONTROL RESULTS

Test Organism	Date Performed	Result
<i>Pseudomonas aeruginosa</i> (ATCC 15442)	4/7/04	1.16×10^5 CFU/carrier
<i>Staphylococcus aureus</i> (ATCC 6538)		1.73×10^5 CFU/carrier
<i>Salmonella choleraesuis</i> (ATCC 10708)		8.8×10^5 CFU/carrier
<i>Salmonella choleraesuis</i> (ATCC 10708)	4/21/04	7.2×10^5 CFU/carrier

CFU = Colony Forming Unit

Project No. A02064

Virox Technologies



Protocol Number: SRC27022404.UJ.1

Page 15 of 16

TABLE 3: NEUTRALIZATION CONFIRMATION CONTROL RESULTS

Test Substance	Test Organism	Date Performed	Inoculum (CFU/mL)	Number of Subculture Tubes Tested	Number of Subculture Tubes Positive
ACCEL TB, Lot 1-3635-REG-US (60 day old)	<i>Pseudomonas aeruginosa</i> (ATCC 15442)	4/7/04	13	6	6
	<i>Staphylococcus aureus</i> (ATCC 6538)		13	6	6
	<i>Salmonella choleraesuis</i> (ATCC 10708)	4/7/04	8	6	6
		4/21/04	26		
ACCEL TB, Lot 2-3646-REG-US	<i>Pseudomonas aeruginosa</i> (ATCC 15442)	4/7/04	13	6	6
	<i>Staphylococcus aureus</i> (ATCC 6538)		13	6	6
	<i>Salmonella choleraesuis</i> (ATCC 10708)	4/7/04	8	6	6
		4/21/04	26		
ACCEL TB, Lot 3-3647-REG-US	<i>Pseudomonas aeruginosa</i> (ATCC 15442)	4/7/04	13	6	6
	<i>Staphylococcus aureus</i> (ATCC 6538)		13	6	6
	<i>Salmonella choleraesuis</i> (ATCC 10708)	4/7/04	8	6	6
		4/21/04	26		

CFU = Colony Forming Unit

Project No. A02064

Virox Technologies



Protocol Number: SRC27022404.UD.1

Page 16 of 16

TABLE 4: TEST RESULTS

Test Substance	Test Organism	Date Performed	Sample Dilution*	Number of Carriers		
				Exposed	Showing Growth**	
ACCEL TB, Lot 1-3635-REG-US (60 day old)	<i>Pseudomonas aeruginosa</i> (ATCC 15442)	4/7/04	RTU	1°=60	1°=0	
	<i>Staphylococcus aureus</i> (ATCC 6538)			2°=60	2°=0	
	<i>Salmonella choleraesuis</i> (ATCC 10708)	4/7/04		1°=60	1°=0	
		4/21/04		2°=60	2°=1	
	ACCEL TB, Lot 2-3646-REG-US	<i>Pseudomonas aeruginosa</i> (ATCC 15442)		4/7/04	1°=60	1°=0
		<i>Staphylococcus aureus</i> (ATCC 6538)			2°=60	2°=1
<i>Salmonella choleraesuis</i> (ATCC 10708)		4/7/04	1°=60	1°=0		
		4/21/04	2°=60	2°=3		
ACCEL TB, Lot 3-3647-REG-US		<i>Pseudomonas aeruginosa</i> (ATCC 15442)	4/7/04	1°=60	1°=0	
		<i>Staphylococcus aureus</i> (ATCC 6538)		2°=60	2°=0	
	<i>Salmonella choleraesuis</i> (ATCC 10708)	4/7/04	1°=60	1°=0		
		4/21/04	2°=60	2°=0		

* RTU = Ready to use

** Number of carriers showing growth of the test organism.

1° Primary Subculture

2° Secondary Subculture